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Note

Rapid determination of aflatoxin M1 in milk and dairy products by high-performance liquid chromatography

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Aflatoxin M1 occurs as a metabolite of aflatoxin B1 in milk from cows fed on mould-contaminated feed. Two techniques are available for the determination of aflatoxin M1: thin-layer chromatography with direct semi-quantitative visual determination and high-performance liquid chromatography (HPLC) with quantitative spectrofluorimetric detection.

As the allowable level of aflatoxin M1 in milk, which was established as 500 ng/l (0.5 ppb) by the U.S. Food and Drugs Administration in $1977^{1,2}$, has decreased to 50 ng/l for milk and 10 ng/l for dairy babyfoods in Switzerland³ and around 30 ng/l in the EEC countries⁴, the above techniques are becoming unsuitable. However, when working on pure aflatoxin M1 solutions, HPLC has sufficient sensitivity.

In order to determine aflatoxin M1 in milk by HPLC, a preliminary extraction and clean-up procedure was developed after comparison with previous methods, including purification on a column of silica^{1,5-9}, alumina⁷, Sep-Pak¹⁰, C₁₈ Sep-Pak¹¹,Celite⁷, Hyflo Supercel¹⁰ or cellulose¹², precipitation with lead salts^{6,12-14}, cadmium salts⁸ or sodium chloride¹, delipidation with a low-polarity solvent such as hexane¹² or diethyl ether¹⁴ and extraction with chloroform¹², acetone^{5,6,13}, methanol-chloroform¹⁵, or isopropanol-chloroform⁷. These techniques generally involve injection of the concentrated extract into a silica HPLC column¹⁵ or, more often, a C₁₈ grafted silica column^{6,10,11,13}. Elution is usually achieved with either acetonitrile-water⁹⁻¹¹ or acetonitrile-methanol-water¹³ and is followed by spectrofluorimetric detection^{6,10,11,13,15}.

The procedure presented here uses zinc hydroxide precipitation, followed by hexane clean-up. A subsequent vacuum concentrated, chloroform extract is injected on to a C_{18} grafted silica column and eluted with acetonitrile-methanol-water.

EXPERIMENTAL

Apparatus

A Kontron liquid chromatograph with an LC 410 pump and an SFM 23 fluorescence detector with $10-\mu$ l cuvette, operated at 365 nm (excitation) and with a 385 nm cut-off detection filter, a Rheodyne injector with a 20- μ l loop and a Li-Chrosorb RP-18 (5 μ m)(Merck, Darmstadt, G.F.R.), 25 cm × 4 mm I.D. column were used. Other apparatus consisted of a rotary vacuum evaporator, a water-bath and an ILA CX blender (20,000 rpm).

Reagents

Analytical-reagent grade sodium sulphate, sodium hydroxide (8% solution), zinc sulphate (28% solution), chloroform, methanol, hexane and acetonitrile were used.

Aflatoxin M1 (Sigma, St. Louis, MO, U.S.A.) was dissolved in chloroform to give a 10 μ g/ml standard solution.

Procedure

The whole procedure described below requires 30 min.

Sample preparation: A 20-ml volume of liquid milk (or 2 g powdered milk) was diluted to 50 ml with doubly distilled water, or 2 g of cheese, was thoroughly blended with 50 ml of doubly distilled water at 40°C.

Deproteinization. To the above samples were added 14 ml of 28% zinc sulphate solution and 8 ml of 8% sodium hydroxide solution and the mixture was shaken gently and sieved through Whatman No. 1 filter-paper on a büchner funnel.

Purification. The filtrate, shaken twice with 50 ml of chloroform for 1 min in a blender, was poured into a separatory funnel. The chloroform extracts were dried over sodium sulphate and concentrated first in a rotary vacuum evaporator and then under a nitrogen flow in a 3-ml septum vial on a water-bath. The dry residue was dissolved into 100 μ l of methanol.

HPLC. A 20- μ l volume of the above methanol solution was injected on to the column and eluted with a mobile phase consisting of acetonitrile-methanol-water (20:5:75) at a flow-rate of 1 ml/min.

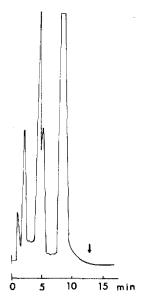
RESULTS AND DISCUSSION

Uncontaminated liquid milk samples were spiked with aflatoxin M1 using five samples per dose level. The recoveries are shown in Table I.

TABLE I RECOVERY OF AFLATOXIN M1 FROM MILK

Aflatoxin MI added (ng/l)	Recovery \pm standard deviation (%)
10	89.8 ± 10.2
50	80.2 ± 2.8
100	83 ± 3.4

The method is accurate and reliable, giving a recovery of ca. 80% at the low ppb level. The clean-up step with zinc hydroxide yields a clear filtrate and no emulsion with hexane or chloroform. A slight modification of Gregory and Manley's mobile phase¹³ allows for slight retardation of the aflatoxin M1 peak, with good separation from impurities (Figs. 1 and 2).



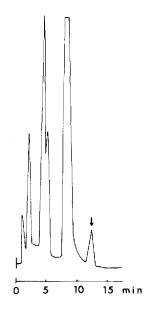


Fig. 1. Chromatogram of an aflatoxin M1-free milk.

Fig. 2. Chromatogram of an aflatoxin M1-fortified milk at 0.025 ppb.

CONCLUSION

The HPLC method described here allows, through a very efficient clean-up of the dairy sample, the rapid determination of aflatoxin M1 with a detection limit of 10 ng/l.

REFERENCES

- I R. D. Stubblefield, J. Amer. Oil Chem. Soc., 56 (1979) 800.
- 2 L. Stoloff, J. Food Protect., 43 (1980) 226.
- 3 F. Y. Tripet, C. Riva and J. Vogel, Lait, 61 (1981) 634.
- 4 D. S. P. Patterson, E. M. Glancy and B. A. Roberts, Food Cosmet. Toxicol., 18 (1980) 35.
- 5 M. Fukayama, W. Winterlin and D. P. H. Sieh. J. Ass. Offic. Anal. Chem., 63 (1980) 927.
- 6 R. M. Beebe and D. M. Takayashi, J. Agr. Food Chem., 28 (1980) 481.
- 7 P. Lafont, M. Siriwardana, J. Jacquet, M. Gaillardin and J. Sarfati, Lait, 61 (1981) 275.
- 8 L. G. M. T. Tuinstra and J. M. Bronsgeest, J. Chromatogr., 111 (1975) 448.
- 9 R. Gauch, U. Leuenberger and E. Baumgartner, J. Chromatogr., 178 (1979) 543.
- 10 J. M. Fremy, T. Cariou and C. Terrier, Ann. Falsif. Expert. Chim., 74 (1981) 547.
- 11 W. Winterlin, G. Hall and D. P. H. Hsieh, Anal. Chem., 51 (1979) 1873.
- 12 W. A. Pons, A. F. Cucullu and L. S. Lee, J. Ass. Offic. Anal. Chem., 56 (1973) 1431.
- 13 J. F. Gregory and D. Manley, J. Ass. Offic. Anal. Chem., 64 (1981) 144.
- 14 R. D. Stubblefield, J. Ass. Offic. Anal. Chem., 57 (1974) 847.
- 15 M. Blanc, Ann. Falsif. Expert. Chim., 72 (1979) 427.